Supporting Information

Dramatic Enhancement of Antagonistic Activity on Vitamin D Receptor: A Double Functionalization of 1α -Hydroxyvitamin D_3 - 26,23-Lactones

Nozomi Saito,[†] Hiroshi Saito,[‡] Miyuki Anzai,[‡] Akihiro Yoshida,^{†,§} Toshie Fujishima,[†] Kazuya Takenouchi, Daishiro Miura,[‡] Seiichi Ishizuka,[‡] Hiroaki Takayama,^{†,¶} and Atsushi Kittaka^{†,*}

[†]Faculty of Pharmaceutical Sciences, Teikyo University, Kanagawa 199-0195, Japan [‡]Teijin Institute for Bio-Medical Research, Tokyo 191-8512, Japan

p 1 ~ p 13 : experimental procedure

p 14 : charts of VDR binding assay

p $15 \sim p 16$: charts of HL-60 cell differentiation

General

All manipulations were performed under an argon atmosphere unless otherwise mentioned. All solvents and reagents were purified when necessary using standard procedures. Column chromatography was performed on silica gel 60 N (Kanto Chemical CO., Inc., $100-210~\mu m$), and flash column chromatography was performed on silica gel 60 (Merck, $40-63~\mu m$).

Experimental Section

Synthesis of Aldehyde <u>8</u>

The aldehyde **8** was synthesized from known compound **18** (Scheme 6).

Compound 19: To a suspension of bromomethyltriphenylphosphonium bromide (5.9 g, 14 mmol) in THF (20 mL) was added a solution of NaHMDS in THF (1.0 M, 14 mL, 14 mmol) at 0 °C, and the mixture was stirred at the same temperature for 1 h. To the mixture was added a solution of 18 (Hijikuro, I.; Doi, T.; Takahashi, T. J. Am. Chem. Soc. 2001, 123, 3716) (1.0 g, 2.7 mmol) in THF (20 mL) at 0 °C, and the resulting mixture was stirred at the same temperature for 2 h. To the mixture was added a saturated NH₄Cl aq. solution at 0 °C, and the aqueous layer was extracted with AcOEt. The organic layer was washed with a saturated NaCl aq. solution, dried over Na₂SO₄, and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 20/1) to give **19** (920 mg, 76%) as a colorless oil. $[\alpha]_D^{26}$ +72.4 (c 1.38, CHCl₃); IR (neat) 1647, 1599, 1360, 1176 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.53 (s, 3 H), 1.00 (d, J = 6.6Hz, 3 H), 1.05-1.81 (m, 10 H), 1.88-2.00 (m, 2 H), 2.45 (s, 3 H), 2.91 (m, 1 H), 3.82 (dd, J = 9.3, 6.1 Hz, 1 H), 3.96 (dd, J = 9.3, 3.2 Hz, 1 H), 5.64 (s, 1 H), 7.34 (d, J = 8.1 Hz, 2 H), 7.78 (d, J = 8.1 Hz, 2 H)Hz, 2 H); ¹³C NMR (150 MHz, CDCl₃) δ 11.8, 17.0, 21.6, 22.0, 22.4, 36.8, 39.5, 45.4, 51.4, 55.4, 75.3, 97.3, 127.9, 129.8, 133.1, 144.5, 144.6; EI-LRMS *m/z* 440 (M⁺), 361, 268, 227, 189, 172, 91; EI-HRMS calcd for $C_{21}H_{29}O_3^{79}Br$ 440.1021, found 440.1023.

Compound 8: To a solution of **19** (1.2 g, 2.8 mmol) in DMSO (3 mL) was added KCN (363 mg, 5.6 mmol), and the mixture was stirred at 70 °C for 1.5 h. The mixture was diluted with Et_2O , and the organic layer was washed with H_2O and saturated NaCl aq. solution, dried over Na_2SO_4 , and concentrated. The residue was dissolved in CH_2Cl_2 (5.5 mL). To the solution was added a

solution of DIBAL-H in toluene (1.0 M, 3 mL, 3.1 mmol) at 0 °C, and the mixture was stirred at the same temperature for 1.5 h. To the mixture was added, 10% potassium sodium tartrate aq. solution, and the aqueous layer was extracted with Et₂O. The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄ and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/Et₂O = 30/1) to give **8** (692 mg, 2.3 mmol in 2 steps) as a colorless oil. $[\alpha]_D^{20}$ +86.1 (c 1.08, CHCl₃); IR (neat) 2950, 1725, 1381 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.62 (s, 3 H), 1.04 (d, J = 6.6 Hz, 3 H), 1.20-1.40 (m, 3 H), 1.45-1.75 (m, 5 H), 1.90 (m, 1 H), 1.95-2.15 (m, 3 H), 2.20 (ddd, J = 16.0, 9.3, 3.2 Hz, 1 H), 2.47 (dd, J = 16.0, 2.7 Hz, 1 H), 2.89 (m, 1 H), 5.67 (s, 1 H), 9.76 (dd, J = 3.2, 1.2 Hz, 1 H); ¹³C NMR (150 MHz, CDCl₃) δ 11.8, 20.0, 21.9, 22.4, 27.7, 30.9, 31.7, 39.6, 45.5, 50.7, 55.4, 55.7, 97.7, 144.6, 203.0; EI-LRMS m/z 298 (M⁺) 254, 227, 148; EI-HRMS calcd for C₁₅H₂₃O⁷⁹Br 298.0932, found 298.0934.

Synthesis of syn-lactones <u>10</u> and <u>11</u>.

syn-Lactones 10 and 11: To a suspension of CrCl₃ (811 mg, 5.1 mmol) in THF (26 mL) was added LiAlH₄ (97 mg, 2.6 mmol) at 0 °C, and the mixture was stirred at room temperature for 30 min. To the mixture were added a solution of 9 (494 mg, 2.6 mmol) in THF (8 mL) and a solution of 8 (385 mg, 1.3 mmol) at room temperature, and the resulting mixture was stirred at the same temperature for 1 h. To the mixture was added H₂O at 0 °C, and the aqueous layer was extracted with Et₂O. The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄, The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 10/1) to give a mixture of 10 and 11 (467 mg, 95 %, ratio of 1 to 1.2). Further separation was performed by recycle-HPLC (column: SHIMADZU Shim-pack PREP-SIL(H)KIT, eluent: hexane/AcOEt = 3/1, flow rate: 10 mL/min, detector:UV (235 nm)). Spectral data of 10: $[\alpha]_D^{25}$ +41.3 (c 1.09, CHCl₃); IR (neat) 1751, 1662, 1631, 1265, 1163 cm⁻¹; ¹H NMR (600 MHz, $CDCl_3$) δ 0.58 (s, 3 H), 1.06 (d, J = 6.9 Hz, 3 H), 1.14 (d, J = 7.0 Hz, 3 H), 1.22-1.51 (m, 5 H), 1.52-1.72 (m, 6 H), 1.96 (m, 1 H), 1.98-2.05 (m, 2 H), 2.88 (m, 1 H), 3.11 (dddq, J = 2.0, 2.0, 6.8, 7.0 Hz, 1 H), 4.60 (ddd, J = 8.3, 6.8, 5.2 Hz, 1 H), 5.84 (d, J = 2.1 Hz, 1 H), 5.65 (s, 1 H), 6.19 (d, J = 3.3) = 2.1 Hz, 1 H); 13 C NMR (100 MHz, CDCl₃) δ 11.9, 14.6, 19.8, 22.1, 22.6, 27.8, 31.1, 34.4, 36.4, 38.1, 39.8, 45.6, 55.7, 56.1, 80.1, 97.5, 120.4, 141.2, 144.7, 170.1; EI-LRMS m/z 380 (M⁺), 301, 227, 147, 105; EI-HRMS calcd for $C_{20}H_{29}O_2^{79}Br$ 380.1351, found 380.1347. **Spectral data of 11**: $[\alpha]_D^{25}$ +179.4 (c 1.28, CHCl₃); IR (neat) 1765, 1664, 1631, 1267, 1124 cm⁻¹; ¹H NMR (600 MHz, $CDCl_3$) δ 0.59 (s, 3 H), 1.01 (d, J = 6.6 Hz, 3 H), 1.10 (ddd, J = 13.3, 10.8, 1.9 Hz, 1 H), 1.13 (d, J = 13.3) = 7.1 Hz, 3 H, 1.20 - 1.35 (m, 3 H), 1.40 - 1.71 (m, 6 H), 1.75 (m, 1 H), 1.86 (m, 1 H), 1.97 (ddd, J = 1.00 - 1.00 (m, 1 H), 1.00 (ddd, J = 1.00 - 1.00 (m, 1 H), 1.00 (ddd, J = 1.00 - 1.00 (m, 1 H), 1.00 (ddd, J = 1.00 - 1.00 (m, 1 H), 1.00 (ddd, J = 1.00 - 1.00 (m, 1 H), 1.00 (ddd, J = 1.00 - 1.00 (m, 1 H), 1.00 (ddd, J = 1.00 - 1.00 (ddd, J = 1.00 (ddd, J = 1.00 - 1.00 (ddd, J = 1.00 - 1.00 (ddd,12.4, 6.7, 1.1 Hz, 1 H), 2.03 (br d, J = 12.4 Hz, 1 H), 5.76 (m, 1 H), 3.17 (ddq, J = 2.5, 7.7, 7.1 Hz, 1 H), 4.68 (ddd, J = 11.8, 7.7, 1.9 Hz, 1 H), 5.53 (d, J = 2.8 Hz, 1 H), 5.65 (s, 1 H), 6.22 (d, J = 2.8H ,1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.0,13.9, 18.5, 22.1, 22.6, 27.6, 31.1, 32.5, 36.9, 37.7, 39.9, 45.6, 55.9, 56.3, 78.3, 97.5, 120.5, 140.6, 144.6, 170.1; EI-LRMS m/z 380 (M⁺), 301, 227, 147, 105; EI-HRMS calcd for $C_{20}H_{29}O_2^{79}Br$ 380.1350, found 380.1353.

The relative stereochemistries of **10** and **11** were determined by NOE experiments to be *syn*-orientation, respectively (Figure 4).

Figure 4. NOE NOE NOE NOE NOE H₂₃ to
$$H_{24}$$
 6.1% H_{24} to H_{23} 5.5% Br 10 Br 11

Determination of absolute stereochemistries of C23 and C24 positions on lactone ring.

Transformation of 10 into MTPA ester 20

The lactone derivative 10 was transformed into MTPA esters 20a and 20b (Scheme 7).

Compound 12: To a solution of **10** (17.5 mg, 46 μmol) in toluene (1 mL) was added a solution of DIBAL-H (1.04 M solution in toluene, 0.18 mL, 0.19 mmol) at 0 °C, and the mixture was stirred at room temperature for 2 h. To the mixture was added 10% potassium sodium tartrate aq. solution at 0 °C, and the aqueous layer was extracted with Et₂O. The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄, and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 3/1) to give diol (17 mg, 95%) as a colorless oil. [α]_D²⁷ +74.1 (c 2.28, CHCl₃); IR (neat) 3239, 1631, 1024 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.57 (s, 3 H), 1.00 (d, J = 6.6 Hz, 3 H), 1.02 (d, J = 7.1 Hz, 3 H), 1.15-1.72 (m, 11 H), 1.86-2.06 (m, 3 H), 2.01 (dq, J = 2.1, 7.1 Hz, 1 H), 2.70-3.05 (m, 3 H), 3.56 (ddd, J = 7.4, 6.0, 2.4 Hz, 1 H), 4.04 (dd, J = 12.9, 0.49 Hz, 1 H), 4.13 (dd, J = 12.9, 0.73 Hz, 1 H), 4.96 (s, 1 H), 6.26 (br d, J = 0.98 Hz, 1 H), 5.63 (br s, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 11.4, 12.0, 19.5, 22.2, 22.6, 28.0, 31.1, 34.6, 39.9, 40.4, 41.7, 45.6, 55.8, 56.6, 65.1, 72.1, 97.4, 113.1, 144.8, 151.31; EI-LRMS m/z 384 (M⁺), 298, 254, 227, 175, 147; EI-HRMS calcd for C₂₀H₃₃O₂⁷⁹Br 384.1664, found 384.1664.

To a solution of the diol (220 mg, 0.57 mmol) in CH_2Cl_2 (2.9 mL) were added pyridine (0.19 mL, 2.4 mmol) and PivCl (0.09 mL, 0.73 mmol) at 0 °C, and the mixture was stirred at the same temperature for 7 h. To the mixture was added H_2O , the aqueous layer was extracted with Et_2O . The organic layer was washed with saturated NaCl aq. solution, dried over Na_2SO_4 , and concentrated. The residue was purified by flash column chromatography on silica gel

(hexane/AcOEt = 10/1) to give **12** (226 mg, 84%) as a colorless oil. $[\alpha]_D^{26} + 59.5$ (c 1.25, CHCl₃); IR (neat) 3503, 1730, 1649, 1284, 1153 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.56 (s, 3 H), 1.05 (d, J = 6.6 Hz, 3 H), 1.04 (d, J = 6.9 Hz, 3 H), 1.15-1.80 (m, 12 H), 1.23 (s, 9 H), 1.90-2.10 (m, 3 H), 2.26 (dq, J = 2.8, 6.9 Hz, 1 H), 2.87 (m, 1 H), 3.79 (m, 1 H), 4.52 (d, J = 13.7 Hz, 1 H), 4.59 (d, J = 13.7 Hz, 1 H), 5.02 (s, 1 H), 5.17 (d, J = 1.2 Hz, 1 H), 5.63 (s, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 11.3, 12.0, 19.6, 22.2, 22.6, 27.3 (3 C), 28.0, 31.1, 34.6, 38.9, 39.9, 40.4, 40.7, 45.6, 55.9, 56.9, 66.1, 70.8, 97.4, 112.6, 144.8, 146.9, 177.9; EI-LRMS m/z 468 (M⁺), 389, 299, 269, 227, 170, 147; EI-HRMS calcd for $C_{25}H_{41}O_3^{79}$ Br 468.2239, found 468.2240.

(S)-MTPA ester 20a: To a solution of 12 (16 mg, 33 µmol) in pyridine (1.5 mL) was added (R)-(-)methoxytrifluoromethylphenylacetyl chloride (MTPACl) (10 μL, 53 μmol) at 0 °C, and the mixture was stirred at room temperature for 16 h. To the mixture was added H₂O, the aqueous layer was extracted with Et₂O. The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄, and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 10/1) to give **20a** (22 mg, 97%) as a colorless oil. $[\alpha]_D^{22}$ +22.2 (c 1.69, CHCl₃); IR (neat) 1738, 1730, 1651, 1277, 1155 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 0.55 (s, 3 H), 0.10 (d, J = 6.1 Hz, 3 H), 1.09 (d, J = 6.9 Hz, 3 H), 1.22 (s, 9 H), 1.20-1.40 (m, 4 H), 1.44 (m, 1 H), 1.50-1.61 Hz(m, 3 H), 1.61-1.71 (m, 2 H), 1.76 (ddd, J = 8.9, 8.3, 7.2 Hz, 1 H), 1.88-2.01 (m, 3 H), 2.47 (dq, J = 8.9, 8.3, 7.2 Hz, 1 H)3.0, 6.9 Hz, 1 H), 2.88 (m, 1 H), 3.49 (s, 3 H), 4.55 (d, J = 13.5 Hz, 1 H), 4.62 (d, J = 13.5 Hz, 1 H), 4.96 (s, 1 H), 5.16 (s, 1 H), 5.35 (ddd, J = 8.9, 4.6, 3.0 Hz, 1 H), 5.65 (dd, J = 1.7, 1.7 Hz, 1 H), 7.35-7.42 (m, 3 H), 7.48-7.52 (m, 2 H); ¹³C NMR (150 MHz, CDCl₃) δ 11.8, 12.1, 19.1, 22.0, 22.5, $27.2 (3 C), 27.8, 31.0, 33.9, 37.5, 37.8, 38.8, 39.8, 45.5, 55.3, 55.7, 56.3, 66.3, 76.5, 84.7 (q, {}^{2}J_{C.F} =$ 27.6 Hz), 97.6, 114.5, 122.4 (q, ${}^{1}J_{\text{C-F}} = 288 \text{ Hz}$), 127.7 (2 C), 128.3 (2 C), 129.5, 131.4, 144.8, 145.0, 166.1, 178.0; EI-LRMS m/z 605 (M⁺-Br), 452, 435, 371, 227, 189; EI-HRMS calcd for $C_{35}H_{48}O_5F_3$ (M⁺-Br) 605.3454, found 605.3449.

(*R*)-MTPA ester <u>20b</u>: In a similar manner to that for the synthesis of **20a** from **12**, a crude product, which was obtained from **12** (16 mg, 35 μmol), (*S*)-(+)-MTPACl (10 μL, 53 μmol) in pyridine at 50 °C for 24 h, was purified by flash column chromatography on silica gel (hexane/AcOEt = 10/1) to give **20b** (24 mg, quant) as a colorless oil. $[\alpha]_D^{27}$ +51.4 (*c* 1.72, CHCl₃); IR (neat) 1740, 1655, 1381, 1273, 1020 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 0.55 (s, 3 H), 1.03 (d, *J* = 5.8 Hz, 3 H), 1.07 (d, *J* = 6.9 Hz, 3 H), 1.21 (s, 9 H), 1.25-1.41 (m, 4 H), 1.46 (m, 1 H), 1.52-1.73 (m, 5 H), 1.85 (ddd, *J* = 10.2, 8.9, 8.9 Hz, 1 H), 1.90-2.30 (m, 3 H), 2.46 (dq, *J* = 6.9, 2.8 Hz, 1 H), 2.88 (m, 1 H), 3.52 (s, 3 H), 4.50 (d, *J* = 13.5 Hz, 1 H), 4.55 (d, *J* = 13.6 Hz, 1 H), 5.01 (s, 1 H), 5.34 (ddd, *J* = 8.9, 4.3, 2.8 Hz, 1 H), 5.66 (s, 1 H), 7.35-7.42 (m, 3 H), 7.50-7.54 (m, 2 H); ¹³C NMR (150 MHz, CDCl₃) δ 11.8, 12.0, 19.1, 22.0, 22.5, 27.2 (3 C), 27.9, 31.0, 33.9, 37.3, 38.1, 38.8, 39.8, 45.5, 55.4, 55.7, 56.3, 66.2, 76.2, 84.3 (q, 2 *J*_{C-F} = 27.6 Hz), 97.7, 114.4, 123.4 (q, 1 *J*_{C-F} = 287.9 Hz), 127.4, 128.2 (2 C), 129.5 (2 C), 132.3, 144.5, 144.8, 165.9, 177.3; EI-LRMS *m/z* 605 (M⁺-Br), 452, 435, 371, 227, 189; EI-HRMS calcd for C₃₅H₄₈O₅F₃ 605.3453 (M⁺-Br), found 605.3448.

The values of $\Delta\delta = \delta_{(S)\text{-MTPA ester}}$ - $\delta_{(R)\text{-MTPA ester}}$ in the ¹H NMR spectra of **20** were calculated and shown in Figure 5. These data were considered by applying a modified Mosher's method reported by Kusumi *et al.* (Ohtani, I.; Kusumi, T.; Koshman, Y.; Kakisawa, H. *J. Am. Chem. Soc.* **1991**, *113*, 4092), and the absolute configurations at the C23 position of **20** was determined to be 23*S*. From this result and NOE experiment shown in Figure 4, the absolute stereochemistry at the C24 position

of **20** was determined to be 24*S*.

Figure 5.

$$\Delta \delta = \delta_{(S)\text{-ester}} - \delta_{(R)\text{-ester}}$$

Transformation of 11 into MTPA ester 22

The lactone derivative 11 was transformed into MTPA esters 22a and 22b (Scheme 8).

Compound 21: To a solution of **11** (15 mg, 40 μmol) in toluene (1 mL) was added a solution of DIBAL-H (1.04 M solution in toluene, 0.15 mL, 0.16 mmol) at 0 °C, and the mixture was stirred at room temperature for 2 h. To the mixture was added 10% potassium sodium tartrate aq. solution at 0 °C, and the aqueous layer was extracted with Et₂O. The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄, and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 3/1) to give diol (14 mg, 93%) as an amorphous solid. [α]_D²⁵ +124.1 (*c* 1.12, CHCl₃); IR (nujol) 3250, 1714, 1635 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.59 (s, 3 H), 0.96 (d, *J* = 6.6 Hz, 3 H), 1.03 (m, 1 H), 1.07 (d, *J* = 7.0 Hz, 3 H), 1.20-1.38 (m, 3 H), 1.40-1.73 (m, 7 H), 1.85-2.06 (m, 3 H), 2.30 (dq, *J* = 3.9, 7.0 Hz, 1 H), 2.53 (br s, 2 H), 2.88 (m, 1 H), 3.71 (ddd, *J* = 10.6, 3.9, 1.8 Hz, 1 H), 4.06 (br d, *J* = 12.9, 1 H), 4.13 (br d, *J* = 12.9 Hz, 1 H), 4.93 (s, 1 H), 5.17 (br s, 1 H), 5.64 (s, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.0, 14.5, 18.8, 22.1, 22.6, 27.8, 31.1, 33.0, 39.9, 40.1, 44.3, 45.6, 56.0, 56.4, 65.6, 71.2, 97.4, 112.5, 144.8, 151.0; EI-LRMS m/z 384 (M⁺), 254, 227, 175, 147, 106, 86; EI-HRMS calcd for C₂₀H₃₃O₂⁷⁹Br 384.1664, found 384.1667.

To a solution of the diol (187 mg, 0.49 mmol) in CH_2Cl_2 (3 mL) were added pyridine (0.12 mL, 1.48 mmol) and PivCl (90 μ L, 0.73 mmol) at 0 °C, and the mixture was stirred at the same temperature for 16 h. To the mixture was added H_2O , the aqueous layer was extracted with Et_2O . The organic layer was washed with saturated NaCl aq. solution, dried over Na_2SO_4 , and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 10/1) to give **21** (207 mg, 91%) as a colorless oil. $[\alpha]_D^{25}$ +90.3 (c 1.52, CHCl₃);

IR (neat) 3510, 1730, 1649, 1284, 1153, 1032 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.58 (s, 3 H), 0.95 (d, J = 6.6 Hz, 3 H), 1.02 (m, 1 H), 1.09 (d, J = 6.9 Hz, 3 H), 1.14-1.73 (m, 11 H), 1.22 (s, 9 H), 1.85-2.06 (m, 3 H), 2.15 (dq, J = 5.3, 6.9 Hz, 1 H), 2.86 (m, 1 H), 3.71 (m, 1 H), 4.54 (s, 2 H), 4.97 (s, 1 H), 5.12 (s, 1 H), 5.63 (s, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.1, 14.1, 18.9, 22.2, 22.7, 27.3 (3 C), 27.9, 31.1, 33.1, 38.9, 40.0. 41.1, 43.7, 45.7, 56.0, 56.4, 66.3, 70.3, 97.4, 112.3, 144.8, 146.9, 177.9; EI-LRMS m/z 468 (M⁺), 389, 299, 269, 227, 170, 147; EI-HRMS calcd for $C_{25}H_{41}O_3^{79}$ Br 468.2239, found 468.2234.

(S)-MTPA ester 22a: To a solution of 21 (21 mg, 33 µmol) in pyridine (1.5 mL) were added (R)-(-)-MTPACl (10 µL, 53 µmol) and DMAP (2.8 mg, 23 µmol) at 0 °C, and the mixture was stirred at room temperature for 24 h. To the mixture was added H₂O, the aqueous layer was extracted with The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄, and The residue was purified by flash column chromatography on silica gel concentrated. (hexane/AcOEt = 10/1) to give **22a** (29 mg, 92%) as a colorless oil. $[\alpha]_D^{23}$ +46.6 (c 1.02, CHCl₃); IR (neat) 1738, 1651, 1630, 1381 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 0.47 (s, 3 H), 0.98 (d, J = 6.1Hz, 3 H), 1.05 (d J = 6.9 Hz, 3 H), 1.10 (m, 1 H), 1.20 (m, 1 H), 1.22 (s, 9 H), 1.25 (m,1 H), 1.27 (m, 1 H), 1.40 (m, 1 H), 1.48-1.80 (m, 5 H), 1.78 (ddd, J = 13.7, 11.2, 1.1 Hz, 1 H), 1.87 (m, 1 H),1.92-2.01 (m, 2 H), 2.35 (dq, J = 6.9, 6.9 Hz, 1 H), 2.87 (m, 1 H), 3.51 (s, 3 H), 4.51 (d, J = 13.5 Hz, J = 13.5 Hz1 H), 4.53 (d, J = 13.5 Hz, 1 H), 4.94 (s, 1 H), 5.10 (s. 1 H), 5.34 (ddd, J = 11.2, 6.9, 1.4 Hz, 1 H), 5.64 (br s, 1 H), 7.35-7.42 (m, 2 H), 7.54-7.58 (m, 2 H); 13 C NMR (150 MHz, CDCl₃) δ 11.7, 15.2, 18.5, 22.0, 22.5, 27.2 (3C), 27.6, 30.9, 33.0, 38.8, 39.3, 39.8, 41.4, 45.5, 55.3, 55.8, 56.0, 66.1, 76.3, 84.5 (q, ${}^{2}J_{C-F}$ = 27.6 Hz), 97.6, 113.5, 123.3 (q, ${}^{1}J_{C-F}$ = 289.1 Hz), 127.6, 128.4 (2 C), 129.6 (2 C), 131.9, 144.9, 145.2, 166.3, 177.9; EI-LRMS *m/z* 684 (M⁺), 605, 452, 435, 371, 227, 189; EI-HRMS calcd for C₃₅H₄₈O₅⁷⁹BrF₃ 684.2637, found 684.2645.

(*R*)-MTPA ester <u>22b</u>: In a similar manner to that for the synthesis of **22a** from **21**, a crude product, which was obtained from **21** (20 mg, 35 μmol), (*S*)-(+)-MTPACl (10 μL, 53 μmol) and DMAP (2.6 mg, 21 μmol) in pyridine at room temperature for 24 h, was purified by flash column chromatography on silica gel (hexane/AcOEt = 10/1) to give **22b** (29 mg, quant) as a colorless oil. $[\alpha]_D^{23}$ +79.6 (*c* 2.14, CHCl₃); IR (neat) 1738, 1649, 1630, 1381 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 0.40 (s, 3 H), 0.96 (d, *J* = 6.1 Hz, 3 H), 1.07 (d *J* = 7.2 Hz, 3 H), 1.16 (m, 1 H), 1.20 (m, 1 H), 1.23 (s, 9 H), 1.26 (m,1 H), 1.36 (m, 1 H), 1.45-1.70 (m, 5 H), 1,74 (dd, *J* = 12.9, 10.7 Hz, 1 H), 1.82 (m, 1 H), 1.90-2.00 (m, 2 H), 2.37 (dq, *J* = 6.9, 6.3 Hz, 1 H), 2.86 (m, 1 H), 3.50 (s, 3 H), 4.52 (d, *J* = 14.0 Hz, 1 H), 4.56 (d, *J* = 14.0 Hz, 1 H), 4.96 (s, 1 H), 5.13 (s. 1 H), 5.38 (ddd, *J* = 10.7, 6.3, 1.4 Hz, 1 H), 5.62 (br s, 1 H), 7.35-7.42 (m, 2 H), 7.54-7.58 (m, 2 H); ¹³C NMR (150 MHz, CDCl₃) δ 11.7, ;15.3, 18.5, 22.0, 22.5, 27.2 (3 C), 27.5, 30.9, 32.7, 38.8, 39.3, 39.8, 41.5, 45.5, 55.2, 55.8, 56.0, 66.1, 76.1, 84.6 (q, ²*J*_{C-F} = 27.6 Hz), 97.5, 113.6, 123.4 (q, ¹*J*_{C-F} = 289.1 Hz), 127.6, 128.4 (2 C), 129.6 (2 C), 131.6, 144.9, 145.3, 166.3, 177.9; EI-LRMS *m*/*z* 684 (M⁺), 605, 452, 435, 371, 227, 189; EI-HRMS calcd for C₃₅H₄₈O₅⁷⁹BrF₃ 684.2637, found 684.2629.

The values of $\Delta\delta = \delta_{(S)\text{-MTPA ester}}$ - $\delta_{(R)\text{-MTPA ester}}$ in the ¹H NMR spectra of **22** were calculated and shown in Figure 6. These data were considered by applying a modified Mosher's method, and the absolute configurations at the C23 positions of **22** were determined to be 23*R*. From this result and NOE experiment shown in Figure 4, the absolute stereochemistry at the C24 position of **22** was determined to be 24*R*.

Figure 6.

-0.025

+0.03
+0.042
+0.041
-0.027

OPiv

O-0.034
O-0.034
O-0.034
O-0.034

MTPA

MTPA plane

$$\Delta \delta = \delta(S) \cdot \text{ester} = \delta(R) \cdot \text{ester}$$

 $\Delta \delta = \delta_{(S)\text{-ester}} - \delta_{(R)\text{-ester}}$

Synthesis of anti-lactones 14 and 16.

Compound 13: To a solution of 12 (210 mg, 0.45 mmol) in CH₂Cl₂ (2.2 mL) were added tetrapropylammonium perruthenate (TPAP, 16 mg, 46 µmol) and N-methylmorpholine N-oxide (NMO, 79 mg, 0.67 mmol) at room temperature, and the mixture was stirred at the same temperature for 4 h. After the mixture was filtered through silica gel short column (Et₂O), the filtrate was concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 20/1) to give ketone (195.6 mg, 94%) as colorless oil. $[\alpha]_{D}^{24} + 124.3$ (c 2.93, CHCl₃); IR (neat) 1732, 1649, 1631, 1280, 1147 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.59 (s, 3 H), 0.92 (d, J = 6.4 Hz, 3 H), 1.20 (d, J = 7.1 Hz, 3 H), 1.22 (s, 9 H), 1.29 (m, 1 H), 1.35-1.75 (m, 7 H),1.83 (m, 1 H), 1.93-2.10 (m, 3 H), 2.26 (dd, J = 16.4, 9.9 Hz, 1 H), 2.52 (dd, J = 16.4, 2.8 Hz, 1 H), 2.88 (m, 1 H), 3.18 (q, J = 7.1 Hz, 1 H), 4.53 (s, 2 H), 5.06 (s, 1 H), 5.21 (s, 1 H), 5.64 (s, 1 H); 13 C NMR (100 MHz, CDCl₃) δ 12.0, 15.3, 20.1, 22.1, 22.6, 27.3 (3 C), 27.7, 31.0, 32.8, 38.8, 39.8, 45.6, 47.5, 51.5, 55.6, 58.9, 65.8, 97.5, 114.9, 143.0, 144.6, 177.6, 209.7; EI-LRMS m/z 466 (M⁺), 387, 366, 279, 237, 175; EI-HRMS calcd for $C_{25}H_{39}^{79}BrO_3$ 466.2083, found 466.2083.

To a solution of the above ketone (34 mg, 71.7 µmol) in THF (1 mL) was added LiAlH(O^tBu)₃ (1.0 M solution in THF, 0.72 mL, 0.72 mmol) at 0 °C, and the mixture was stirred at the same temperature for 9 h. To the mixture was added saturated NH₄Cl aq. solution, and the aqueous layer was extracted with AcOEt. The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄, and concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 10/1) to give **13** (31 mg, 91%) as a colorless oil. $[\alpha]_D^{23}$ +71.1 (c 2.06, CHCl₃); IR (neat) 3514, 1728, 1649, 1286, 1153 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.59 (s, 3 H), 0.96 (d, J = 6.6 Hz, 3 H), 1.03 (d, J = 7.1 Hz, 3 H), 1.16 (m, 1 H), 1.22 (s, 9 H), 1.20-1.80 (m, 10 H),1.91 (m, 1 H), 1.98 (ddd, J = 12.4, 6.8, 1.5 Hz, 1 H), 2.03 (m, 1 H), 2.16 (m, 1 H), 2.21 (br s, 1 H), 2.87 (m, 1 H), 3.59 (m, 1 H), 4.50 (d, J = 13.9 Hz, 1 H), 4.58 (d, J = 13.9 Hz, 1 H), 5.04 (s, 1 H),5.11 (d, J = 1.2 Hz, 1 H), 5.63 (s, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.1, 16.3, 18.8, 22.2, 22.7, 27.3 (3 C), 27.8, 31.1, 32.8, 38.9, 40.0, 41.0, 45.7, 46.1, 56.0, 56.3, 65.3, 71.1, 97.4, 113.2, 144.9, 146.3, 178.1; EI-LRMS m/z 468 (M⁺), 390, 229, 178, 68, 57; EI-HRMS calcd for $C_{25}H_{41}^{79}BrO_{3}$ 468.2239, found 468.2243.

tanti-Lactone 14: To a solution of 13 (31 mg, 65 µmol) in toluene (1 mL) was added DIBAL-H (1.04 M solution in toluene, 0.31 mL, 0.32 mmol) at 0 °C, and the mixture was stirred at room temperature for 1 h. To the mixture was added 10% potassium sodium tartrate aq. solution at 0 °C, and the aqueous layer was extracted with AcOEt. The organic layer was washed with saturated

NaCl aq. solution, dried over Na₂SO₄, and concentrated. The residue was dissolved in CH₂Cl₂ (2 mL). To the solution was added MnO₂ (113 mg, 1.3 mmol), and the mixture was stirred at room temperature for 3 days. After the mixture was filtered through silica gel short column (Et₂O), the filtrate was concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 10/1) to give **14** (10 mg, 81% in 2 steps) as a colorless oil. [α]_D¹⁹ +205.8 (c 1.16, CHCl₃); IR (neat) 1759, 1663, 1630, 1314 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.59 (s, 3 H), 1.02 (d, J = 6.6 Hz, 3 H), 1.23 (d, J = 6.6 Hz, 3 H), 1.20-1.95 (m, 12 H), 1.98 (ddd, J = 12.2, 5.4, 1.7 Hz, 1 H), 2.03 (br d, J = 13.2 Hz, 1 H), 2.61 (m, 1 H), 2.89 (m, 1 H), 4.07 (ddd, J = 10.7, 7.3, 2.2 Hz, 1 H), 5.53 (d, J = 3.1 Hz, 1 H), 5.65 (d, J = 1.7 Hz, 1 H), 6.22 (d, J = 3.1 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.1, 16.3, 18.8, 22.2, 22.6, 27.7, 31.1, 33.0, 40.0, 40.9, 41.6, 45.7, 55.9, 56.1, 82.4, 97.6, 120.5, 140.7, 144,6, 170.1; EI-LRMS m/z 380 (M⁺), 301, 227, 147; EI-HRMS calcd for C₂₀H₂₉⁷⁹BrO₂ 380.1351, found 380.1345.

Compound 15: To a solution of **21** (Scheme 8, 273 mg, 0.58 mmol) in CH₂Cl₂ (2.9 mL) were added TPAP (20 mg, 58 μmol) and NMO (102 mg, 87 μmol) at room temperature, and the mixture was stirred at the same temperature for 4 h. After the mixture was filtered through silica gel short column (Et₂O), the filtrate was concentrated. The residue was purified by flash column chromatography on silica gel (hexane/AcOEt = 30/1) to give **15** (252 mg, 93%) as a colorless oil. [α]_D²¹ +25.2 (c 1.35, CHCl₃); IR (neat) 1732, 1716, 1651, 1631, 1371, 1147 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.55 (s, 3 H), 0.85 (d, J = 6.6 Hz, 3 H), 1.17 (d, J =7.1 Hz, 3 H), 1.18 (s, 9 H), 1.19-1.30 (m, 3 H), 1.35-1.70 (m, 5 H), 1.79 (m, 1 H), 1.88-2.05 (m, 3 H), 2.22 (dd, J = 16.7, 9.9 Hz, 1 H), 2.45 (dd, J = 16.7, 2.4 Hz, 1 H), 2.82 (m,1 H), 3.18 (q, J = 7.1 Hz, 1 H), 4.46 (d, J = 14.7 Hz, 1 H), 4.50 (d, J = 14.7 Hz, 1 H), 4.99 (s, 1 H), 5.16 (s, 1 H), 5.59 (s, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 11.9, 15.5, 19.7, 22.0, 22.5, 27.2 (3 C), 27.7, 30.9, 32.2, 38.8, 39.7, 45.5, 47.6, 50.1, 55.5, 55.8, 65.7, 97.4, 114.6, 143.0, 144.5, 177.4, 209.0; EI-LRMS m/z 466 (M⁺), 387, 364, 279, 237, 175, 137; EI-HRMS calcd for C₂₅H₃₉⁷⁹BrO₃ 466.2082, found 466.2086.

anti-Lactone 16: To a solution of 15 (185 mg, 0.40 mmol) in toluene (4 mL) was added DIBAL-H (1.04 M solution in toluene, 1.9 mL, 2.0 mmol) at -78 °C, and the mixture was stirred at the same temperature for 1.5 h. To the mixture was added 10% potassium sodium tartrate aq. solution at 0 °C, and the aqueous layer was extracted with Et₂O. The organic layer was washed with saturated NaCl aq. solution, dried over Na₂SO₄, and concentrated. The residue was dissolved in CH₂Cl₂ (2 mL). To the solution was added MnO₂ (668 mg, 7.7 mmol), and the mixture was stirred at room temperature for 38 h. After the mixture was filtered through silica gel short column (Et₂O), the filtrate was concentrated. The residue was purified by preparative thin-layer chromatography on silica gel (hexane/AcOEt = 10/1) to give 16 (60 mg, 40 % in 2 steps) as a colorless oil. [α]_D²¹ +84.2 (c 0.92, CHCl₃); IR (neat) 1755, 1664, 1628, 1313, 1153; ¹H NMR (400 MHz, CDCl₃) δ 0.58 (s, 3 H), 1.07 (d, J = 6.1 Hz, 3 H), 1.25 (d, J = 6.8 Hz, 3 H), 1.20-1.75 (m, 11 H), 1.90-2.10 (m, 3 H), 2.64 (m, 1 H), 2.88 (m, 1 H), 4.07 (dt, J = 6.3, 5.6 Hz, 1 H), 5.53 (d, J = 3.1 Hz, 1 H), 5.65 (s, 1 H), 6.22 (d, J = 3.1 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.0, 17.3, 19.6, 22.2, 22.6, 28.0, 31.1, 34.5, 39.8, 40.8, 41.5, 45.6, 55.8 (2 C), 84.0, 97.5, 120.7, 140.5, 144.7, 170.1; EI-LRMS m/z 380 (M⁺), 301, 227, 147; EI-HRMS calcd for C₂₀H₂₀O₂⁷⁹Br 380.1351, found 380.1354.

General Procedure for the synthesis of vitamin D_3 -lactones: Method A. To a solution of an Aring precursor (1.3 or 1.5 equiv. to a CD-ring precursor) and the CD-ring precursor in toluene were added Et_3N and $Pd(PPh_3)_4$ (30 mol % to the CD-ring precursor), and the mixture was stirred at 110 °C. After the mixture was filtered through silica gel pad, the filtrate was concentrated. The crude product was dissolved in MeOH. To the solution was added (+)-10-camphorsulfonic acid (CSA) at 0 °C, and the mixture was stirred at room temperature. To the mixture was added saturated NaHCO₃ aq. solution, and the aqueous layer was extracted with AcOEt. The organic layer was washed with saturated. NaCl aq. solution, dried over Na_2SO_4 , and concentrated. The residue was purified by flash column chromatography on silica gel or preparative thin-layer chromatography on silica gel to give the vitamin D_3 -lactone derivative. Further purification for biological assays was conducted by reversed-phase recycle HPLC (YMC-Pack ODS column, 20 X 150 mm, 9.9 mL/min, $CH_3CN:H_2O = 85:15$).

General Procedure for the synthesis of vitamin D_3 -lactones: Method B. To a solution of an Aring precursor (1.5 equiv. to a CD-ring precursor) and the CD-ring precursor in toluene were added Et_3N and $Pd(PPh_3)_4$ (30 mol % to the CD-ring precursor), and the mixture was stirred at 110 °C. After the mixture was filtered through silica gel pad, the filtrate was concentrated. The crude product was dissolved in MeCN (1 mL). To the solution was added 10% solution of conc. HF in MeCN (1 mL) at 0 °C, and the mixture was stirred at room temperature. To the mixture was added saturated NaHCO₃ aq. solution, and the aqueous layer was extracted with AcOEt. The organic layer was washed with saturated. NaCl aq. solution, dried over Na_2SO_4 , and concentrated. The residue was purified by flash column chromatography on silica gel or preparative thin-layer chromatography on silica gel to give the vitamin D_3 -lactone derivative. Further purification for biological assays was conducted by reversed-phase recycle HPLC (YMC-Pack ODS column, 20 X 150 mm, 9.9 mL/min, $CH_3CN:H_2O = 85:15$).

(23S,24S)-25-Dehydro- 1α -hydroxy-24-mehtylvitamin $D_3-26,23$ -lactone (4): According to the General Procedure (Method A), a crude product, which was obtained from 10 (35 mg, 92 µmol), 17 (44 mg, 0.12 mmol), Et₃N (1.5 mL) and Pd(PPh₃)₄ (32 mg, 28 μmol) in toluene (3 mL) at 110 °C for 1.5 h, was treated with CSA (47 mg, 0.20 mmol) in MeOH (3 mL) for 1.5 h. After usual work up, the crude product was purified by flash column chromatography on silica gel (hexane/AcOEt = 1/3) to give 4 (20 mg, 48% in 2 steps) as a colorless oil. UV (EtOH) $\lambda_{max} = 265$ nm; $[\alpha]_D^{18} - 17.0$ (c 0.52, CHCl₃); IR (neat) 3395, 1755, 1638, 1269, 1055 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.56 (s, 3 H), 1.05 (d, J = 6.6 Hz, 3 H), 1.13 (d, J = 7.1 Hz, 3 H), 1.20-1.75 (m, 13 H), 1.87-1.95 (m, 2 H), 1.96-2.08 (m, 3 H), 2.31 (dd, J = 13.4, 6.6 Hz, 1 H), 2.59 (dd, J = 13.4, 3.4 Hz, 1 H), 2.82 (dd, J = 13.4, 3.4 Hz, 1 H)12.5, 4.4 Hz, 1 H), 3.11 (dddq, J = 2.2, 2.2, 6.8, 7.1 Hz, 1 H), 4.22 (m, 1 H), 4.43 (m, 1 H), 4.59 (ddd, J = 8.2, 6.8, 5.3 Hz, 1 H), 4.99 (dd, J = 1.5, 1.5 Hz, 1 H), 5.32 (dd, J = 1.5, 1.5 Hz, 1 H), 5.53(d, J = 2.2 Hz, 1 H), 6.01 (d, J = 11.2 Hz, 1 H), 6.18 (d, J = 2.2 Hz, 1 H), 6.37 (d, J = 11.2 Hz, 1 H);¹³C NMR (100 MHz, CDCl₃) δ 12.1, 14.7, 19.9, 22.4, 23.6, 27.8, 29.1, 34.5, 36.4, 38.2, 40.8, 42.9, 45.3, 46.0, 56.2, 56.9, 66.8, 70.8, 80.3, 111.6, 117.1, 120.4, 124.3, 132.9, 141.3, 142.6, 147.5, 170.2; EI-LRMS m/z 440 (M⁺), 422, 404, 251, 105; EI-HRMS calcd for $C_{28}H_{40}O_4$ 440.2987, found 440.2932.

(23S,24R)-25-Dehydro-1α-hydroxy-24-mehtylvitamin D_3 -26,23-lactone (5): According to the General Procedure (Method A), a crude product, which was obtained from 16 (14 mg, 37 μmol), 17 (18 mg, 48 μmol), Et₃N (1 mL) and Pd(PPh₃)₄ (13 mg, 11 μmol) in toluene (2 mL) at 110 °C for 1.5 h, was treated with CSA (20 mg, 86 μmol) in MeOH (1.5 mL) for 1.5 h. After usual work up, the crude product was purified by flash column chromatography on silica gel (hexane/AcOEt = 1/2) to give 5 (7.8 mg, 48% in 2 steps) as an amorphous solid. UV (EtOH) λ_{max} = 265 nm; [α]_D²³ +19.7 (c 0.30, CHCl₃); IR (neat) 3400, 1759, 1630 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.57 (s, 3 H), 1.06 (d, J = 5.9 Hz, 3 H), 1.28 (d, J = 6.8 Hz, 3 H), 1.25-1.80 (m, 13 H), 1.85-2.10 (m, 5 H), 2.32 (dd, J = 13.6, 6.4 Hz, 1 H), 2.55-2.70 (m, 2 H), 2.83 (m, 1 H), 4.07 (dt, J = 5.9, 6.4 Hz, 1 H), 4.23 (m, 1 H), 4.43 (m, 1 H), 5.00 (s, 1 H), 5.33 (s, 1 H), 5.53 (d, J = 2.9 Hz, 1 H), 6.01 (d, J = 11.1 Hz, 1 H), 6.22 (d, J = 2.9 Hz, 1 H), 6.37 (d, J = 11.1 Hz, 1 H); ¹³C NMR (150 MHz, CDCl₃) δ 12.0, 17.2, 19.5, 22.3, 23.5, 27.9, 29.0, 34.5, 40.4, 40.7, 41.5, 42.9, 15.2, 45.9, 56.2, 56.5, 66.9, 70.8, 84.2, 111.8, 117.2, 120.8, 124.9, 133.0, 140.8, 142.8, 147.6, 170.4; EI-LRMS m/z 440 (M⁺), 422, 404, 251, 105; EI-HRMS calcd for C₂₈H₄₀O₄ 440.2927 found 440.2929.

(23*R*,24*S*)-25-Dehydro-1α-hydroxy-24-mehtylvitamin D_3 -26,23-lactone (6): According to the General Procedure (Method A), a crude product, which was obtained from 14 (19 mg, 49 μmol), 17 (27 mg, 74 μmol), Et₃N (1 mL) and Pd(PPh₃)₄ (17 mg, 15 μmol) in toluene (2 mL) at 110 °C for 1.5 h, was treated with CSA (27 mg, 0.12 mmol) in MeOH (1.5 mL) for 1.5 h. After usual work up, the crude product was purified by flash column chromatography on silica gel (hexane/AcOEt = 1/2) to give 6 (7.8 mg, 52% in 2 steps) as an amorphous solid. UV (EtOH) λ_{max} = 265 nm; [α]_D¹⁸ +81.3 (*c* 0.27, CHCl₃); IR (neat) 3383, 1765, 1643, 1247, 1057 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.57 (s, 3 H), 1.01 (d, *J* = 6.4 Hz, 3 H), 1.22 (d, *J* = 6.8 Hz, 3 H), 1.20-1.38 (m, 4 H), 1.40-2.10 (m, 14 H), 2.31 (dd, *J* = 13.4, 6.4 Hz, 1 H), 2.55-2.65 (m, 2 H), 2.82 (dd, *J* = 12.2, 3.9 Hz, 1 H), 4.07 (ddd, *J* = 10.5, 7.3, 2.0 Hz, 1 H), 4.22 (m, 1 H), 4.42 (dd, *J* = 7.6, 4.4 Hz, 1 H), 4.99 (s, 1 H), 5.32 (dd, *J* = 1.7, 1.4 Hz, 1 H), 5.52 (d, *J* = 2.9 Hz, 1 H), 6.01 (d, *J* = 11.2 Hz, 1 H), 6.21 (d, *J* = 2.9 Hz, 1 H), 6.36 (d, *J* = 11.2 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.2, 16.3, 18.8, 22.4, 23.6, 27.7, 29.1, 33.1, 40.6, 40.9, 41.6, 43.0, 45.3, 46.1, 56.4, 56.9, 66.9, 70.8, 82.5, 111.7, 117.1, 120.4, 124.7, 133.0, 140.8, 142.5, 147.4, 170.2; EI-LRMS *m/z* 440 (M⁺), 422, 404, 251, 105; EI-HRMS calcd for C₂₈H₄₄O₄ 440.2927, found 440.2920

(23*R*,24*R*)-25-Dehydro-1α-hydroxy--24-mehtylvitamin D₃-26,23-lactone ($\underline{7}$): According to the General Procedure (Method A), a crude product, which was obtained from 11 (37 mg, 96 μmol), 17 (46 mg, 0.12 mmol), Et₃N (1.5 mL) and Pd(PPh₃)₄ (33 mg, 29 μmol) in toluene (3 mL) at 110 °C for 1.5 h, was treated with CSA (47 mg, 0.20 mmol) in MeOH (1.5 mL) for 45 min. After usual work up, the crude product was purified by flash column chromatography on silica gel (hexane/AcOEt = 1/2) to give 7 (24 mg, 57% in 2 steps) as an amorphous solid. UV (EtOH) λ_{max} = 265 nm; [α]_D²³ +113.9 (*c* 0.38, CHCl₃); IR (neat) 3420, 1757, 1658 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.57 (s, 3 H), 1.02 (d, J = 6.4 Hz, 3 H), 1.08 (m, 1 H), 1.13 (d, J = 7.3 Hz, 3 H), 1.15-1.35 (m, 3 H), 1.40-2.10 (m, 14 H), 2.31 (dd, J = 13.4, 6.6 Hz, 1 H), 2.59 (dd, J = 13.4, 3.3 Hz, 1 H), 2.83 (dd, J = 12.1, 3.8 Hz, 1 H), 3.16 (dq, J = 7.8, 7.3 Hz, 1 H), 4.23 (m, 1 H), 4.43 (m, 1 H), 4.67 (ddd, J = 11.8, 7.8, 2.0 Hz, 1 H), 4.99 (s, 1 H), 5.33 (s, 1 H), 5.52 (d, J = 2.7 Hz, 1 H), 6.01 (d, J = 11.3 Hz, 1 H), 6.21 (d, J = 2.7 Hz, 1 H), 6.36 (d, J = 11.3 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.2, 13.9, 18.6, 22.4,

23.6, 27.7, 29.1, 32.6, 37.0, 37.8, 40.6, 42.9, 45.3, 46.1, 56.4, 57.1, 66.8, 70.7, 78.4, 111.7, 117.1, 120.5, 124.7, 133.0, 140.7, 142.5, 147.4, 170.2; EI-LRMS m/z 440 (M⁺), 422, 404, 378, 289, 209,105; EI-HRMS calcd for $C_{28}H_{40}O_4$ 440.2927, found 440.2935.

(23*S*,24*S*)-25-Dehydro-2α,24-dimethyl-1α-hydroxyvitamin D₃-26,23-lactone ($\underline{4a}$): According to the General Procedure (Method B), a crude product, which was obtained from **10** (19 mg, 49 μmol), **17a** (28 mg, 74 μmol), Et₃N (1.5 mL) and Pd(PPh₃)₄ (17 mg, 15 μmol) in toluene (3 mL) at 110 °C for 1.5 h, was treated with conc. HF in MeCN for 1.5 h. After usual work up, the crude product was purified by flash column chromatography on silica gel (hexane/AcOEt = 1/1) to give **4a** (15 mg, 68% in 2 steps) as an amorphous solid. UV (EtOH) $\lambda_{\text{max}} = 267$ nm; [α]_D +4.39 (*c* 1.13, CHCl₃); IR (neat) 2281, 1774, 1651, 1603, 1263, 1136 cm⁻¹; H NMR (400 MHz, CDCl₃) δ 0.55 (s, 3 H), 1.05 (d, J = 6.6 Hz, 3 H), 1.08 (d, J = 6.8 Hz, 3 H), 1.13 (d, J = 7.1 Hz, 3 H), 1.20-1.75 (m, 13 H), 1.85-2.10 (m, 4 H), 2.23 (dd, J = 13.6, 7.9 Hz, 1 H), 2.66 (dd, J = 13.6, 4.0 Hz, 1 H), 2.82 (m, 1 H), 3.11 (m, 1 H), 3.84 (m, 1 H), 4.31 (m, 1 H), 4.59 (m, 1 H), 5.00 (d, J = 1.7 Hz, 1 H), 5.27 (br s, 1 H), 5.53 (d, J = 2.3 Hz, 1 H), 6.01 (d, J = 11.4 Hz, 1 H), 6.18 (d, J = 2.3 Hz, 1 H), 6.38 (d, J = 11.4 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.1, 12.7, 14.7, 19.8, 22.4, 23.6, 27.9, 29.1, 34.5, 36.4, 38.2, 40.5, 43.5, 44.3, 46.0, 56.2, 56.9, 71.7, 75.4, 80.3, 113.1, 117.0, 120.4, 124.6, 133.0, 141.2, 143.1, 146.4, 170.2; EI-LRMS m/z 454 (M⁺), 436, 418, 265, 166, 148; EI-HRMS calcd for C₂₉H₄₂O₄ 454.3083, found 454.3095.

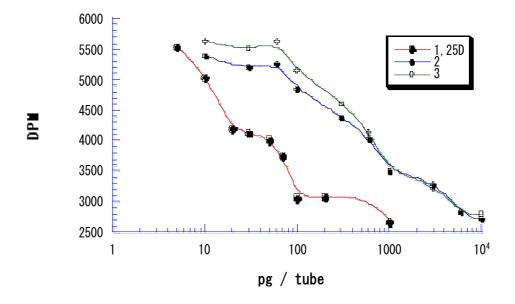
(23S,24R)-25-Dehydro-2α,24-dimethyl-1α-hydroxyvitamin D₃-26,23-lactone ($\underline{5a}$): According to the General Procedure (Method B), a crude product, which was obtained from **16** (12 mg, 31 μmol), **17a** (19 mg, 74 μmol), Et₃N (2 mL) and Pd(PPh₃)₄ (11 mg, 9.3 μmol) in toluene (2 mL) at 110 °C for 1.5 h, was treated with conc. HF in MeCN for 2.5 h. After usual work up, the crude product was purified by column chromatography on silica gel (hexane/AcOEt = 1/1) to give **5a** (10 mg, 71% in 2 steps) as an amorphous solid. UV (EtOH) $\lambda_{\text{max}} = 267$ nm; [α]_D²¹ +45.0 (c 0.64, CHCl₃); IR (neat) 350, 1761, 1667, 1644, 1248, 1066 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.56 (s, 3 H), 1.06 (d, J = 6.8 Hz, 3 H), 1.08 (d, J = 7.3 Hz, 3 H), 1.25 (d, J = 6.8 Hz, 3 H), 1.30-1.75 (m, 14 H), 1.85-2.10 (m, 3 H), 2.23 (dd, J = 13.4, 8.1 Hz, 1 H), 2.55-2.75 (m, 2 H), 2.82 (m, 1 H), 3.83 (m, 1 H), 4.07 (m, 1 H), 4.31 (m, 1 H), 5.00 (s, 1 H), 5.27 (s, 1 H), 5.53 (d, J = 2.9 Hz, 1 H), 6.00 (d, J = 11.2 Hz, 1 H), 6.22 (d, J = 2.9 Hz, 1 H), 6.38 (d, J = 11.2 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 12.2, 12.7, 17.4, 19.7, 22.4, 23.6, 28.1, 29.1, 34.6, 40.5, 40.8, 41.6, 43.5, 44.3, 46.0, 56.2, 56.6, 71.7, 75.4, 84.1, 113.1, 117.0, 120.6, 124.6, 133.0, 140.6, 142.7, 146.4, 170.1; EI-LRMS m/z 454 (M⁺), 436, 418, 265, 166, 148; EI-HRMS calcd for C₂₉H₄₂O₄ 454.3083, found 454.3083.

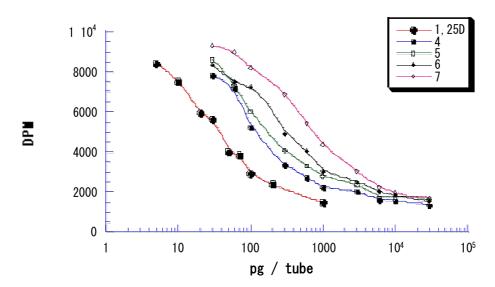
Vitamin D receptor (VDR) binding assay: [26,27-methyl- 3 H]-1α,25-dihydroxyvitamin D₃ (specific activity 6.623 TBq/mmol, 15,000 dpm, 15.7 pg) and various amounts of 1α,25-dihydroxyvitamin D₃ and an analogue to be tested were dissolved in 50 mL of absolute ethanol in 12 x 75-mm polypropylene tubes. 0.2 mg of the chick intestinal VDR and 1 mg of gelatin in 1 mL of phosphate buffer solution (25 nM KH₂PO₄, 0.1 M KCl, 1 mM dithiothreitol, pH 7.4) were added to each tube in an ice bath. The assay tubes were incubated in shaking water bath for 1 h at 25 °C and then chilled in an ice bath. 1 mL of 40% polypropylene glycol 6000 in distilled water was added to each tube, which was the mixed vigorously and centrifuged at 2,260 x g for 60 min at 4 °C.

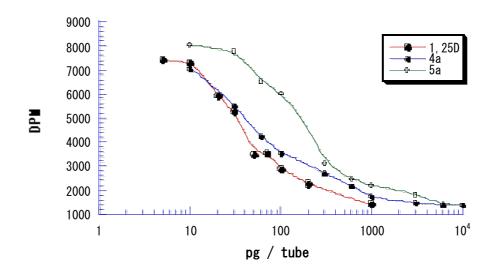
After the supernatant was decanted, the bottom of the tube containing the pellet was cut off into a scintillation vial containing 10 ml of dioxane-based scintillation fluid and the radioactivity was counted with a Beckman liquid scintillation counter (Model LS6500). The relative potency of the analogues were calculated from their concentration needed to displace 50% of [26,27-methyl- 3 H]-1 α ,25-dihydroxyvitamin D_3 from the receptor compared with the activity of 1 α ,25-dihydroxyvitamin D_3 (assigned a 100% value).

Assay for HL-60 cell differentiation: Nitro blue tetrazolium (NBT)-reducing activity was used as a cell differentiation marker. HL-60 cells were cultured in RPMI-1640 medium supplemented with 10% heat-inactivated FCS. Exponentially proliferating cells were collected, suspended in fresh medium and seeded in culture plates (Falcon, Becton Dickinson and Company, Franklin Lakes, NJ). Cell concentration at seeding was adjusted to 2 x 10⁴ cells/mL and the seeding volume was 1 mL/well. An ethanol solution of 1α,25-dihydroxyvitamin D₃ (final concentration: 10⁻⁸ M) and an analogue (final concentration: 10⁻¹¹ to 10⁻⁶ M) was added to the culture medium at 0.1% volume and culture was continued for 96 h at 37 °C in a humidified atmosphere of 5% CO₂/air without medium change. The same amount of vehicle was added to the control culture. NBT-reducing assay was performed according to the method of Collins.¹⁹ Briefly, cells were collected, washed with PBS, and suspended in serum-free medium. NBT/TPA solution (dissolved in PBS) was added. Final concentrations of NBT and TPA were 0.1% and 100 ng/mL, respectively. Then, the cell suspensions were incubated at 37 °C for 25 min. After incubation, cells were collected by centrifugation and resuspended in FCS. Cytospin smears were prepared, and the counter-staining of nuclei was done with Kemechrot solution. At least 500 cells per preparation were observed.

Charts of VDR binding affinity







Charts of HL-60 cell differentiation

